ZipChip CE-ESI-MS for Fast and Efficient Biopharmaceutical Characterization

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Overview

ZipChips are glass microfluidic devices that integrate a CE separation with electrospray ionization performed directly off of the corner of the device. Microfluidic integration of components enables extremely fast and efficient separations, while also making the system small, simple and easy to use. The ZipChip system has previously been available only on Thermo mass spectrometers, but we have recently completed development of a ZipChip interface that is compatible with Sciex mass specs. This presentation provides a demonstration of the capabilities of the ZipChip system when used in combination with a Sciex 6600 TripleTOF for applications relevant to biopharmaceutical labs.

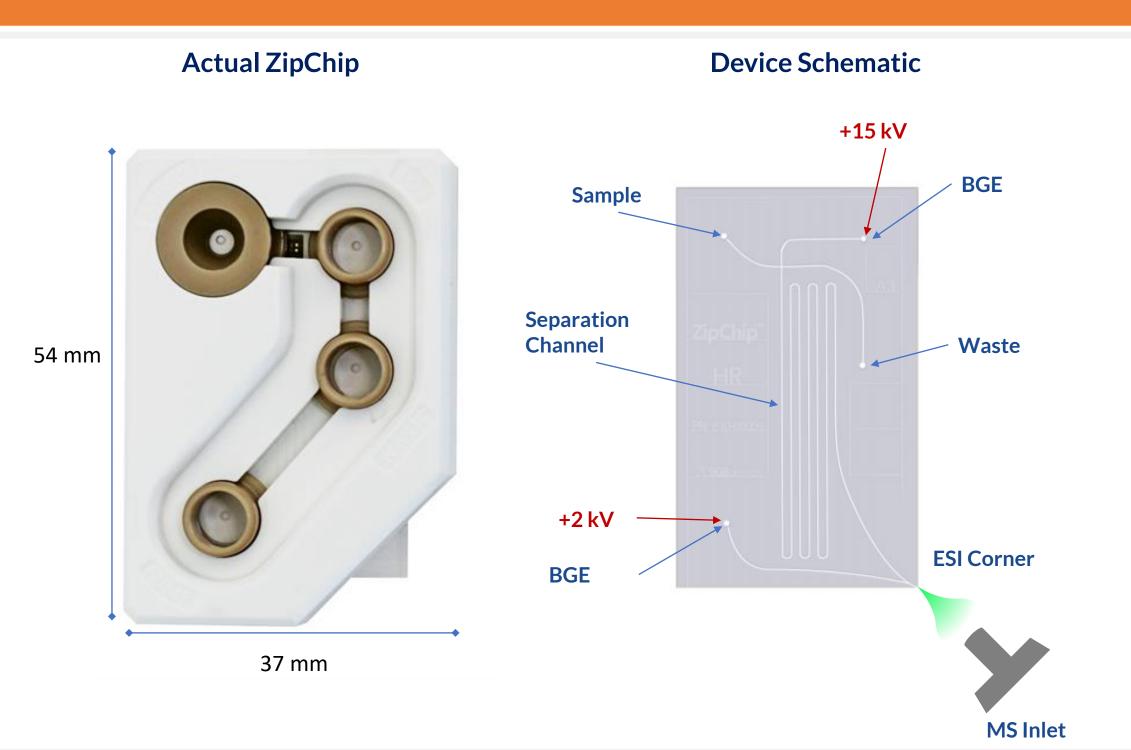




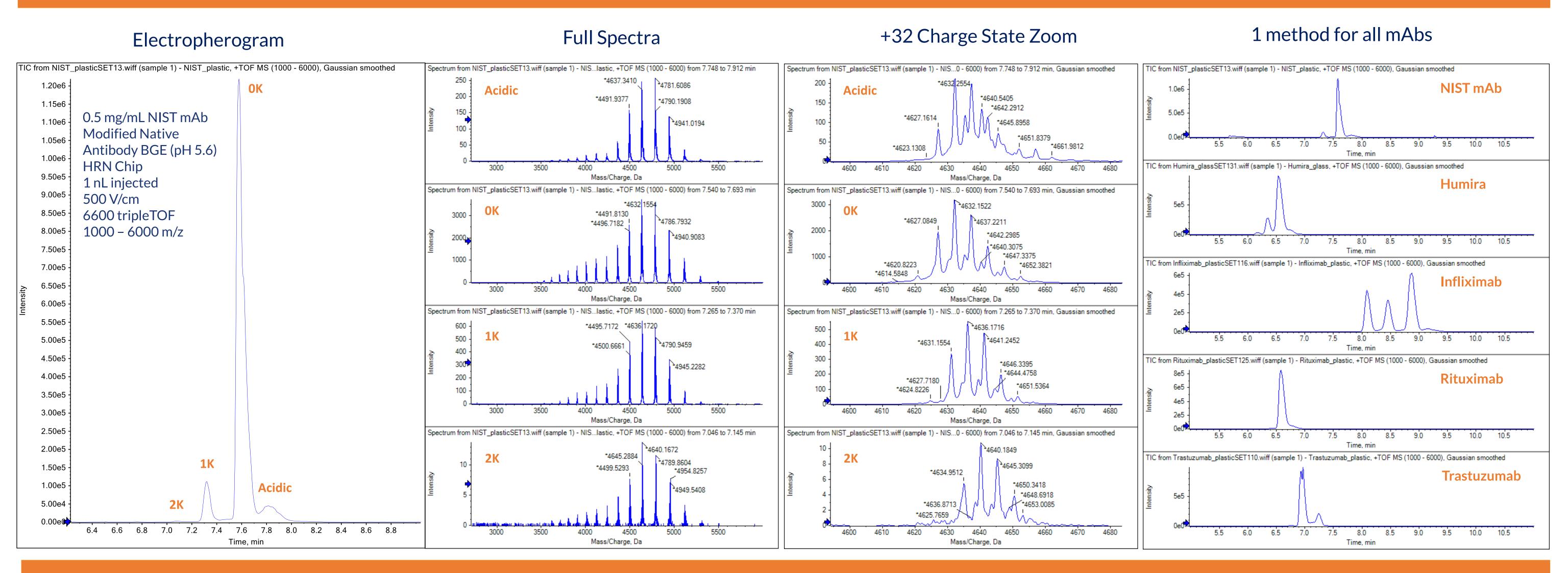
Methods

Instrumentation: All of the data on this poster was acquired using the new ZipChip Sciex Interface (ZC-SI, 908 Devices) mounted on a Sciex 6600 tripleTOF mass spectrometer. ZipChips are available with two different channel lengths; HS chips have a 10-cm long separation channel, while HR chips have a 22-cm long separation channel. All of the work demonstrated here utilized HR chips. All ZipChips utilize a covalently attached, neutral polymer surface coating to prevent analyte interactions and suppress electroosmotic flow. The charge variant work shown here, used a "high resolution native" (HRN) chip. This chip uses a new surface coating process to achieve high resolution protein separations under native conditions.

Sample Prep: Intact mAbs were diluted with background electrolyte (BGE) directly from their formulations to a concentration of 0.5 mg/mL. The NIST mAb subunit sample was digested with the IdeS enzyme and reduced with 100 mM DTT to yield Fc, Fd, and light chain subunits. The peptide mapping sample was digested with Trypsin using the Thermo Smart digest protocol.



Native Antibody Charge Variant Analysis with ZipChip CE-ESI-MS



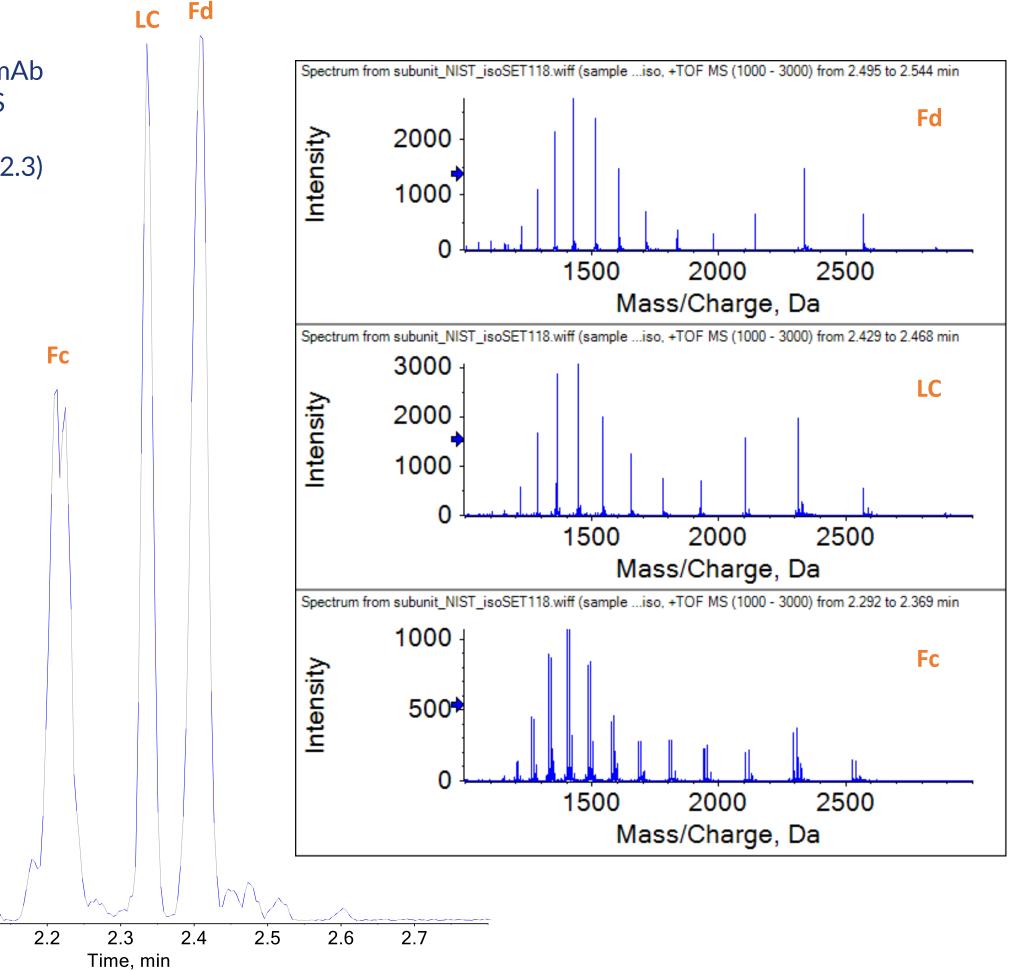
Multilevel Characterization

Intensity

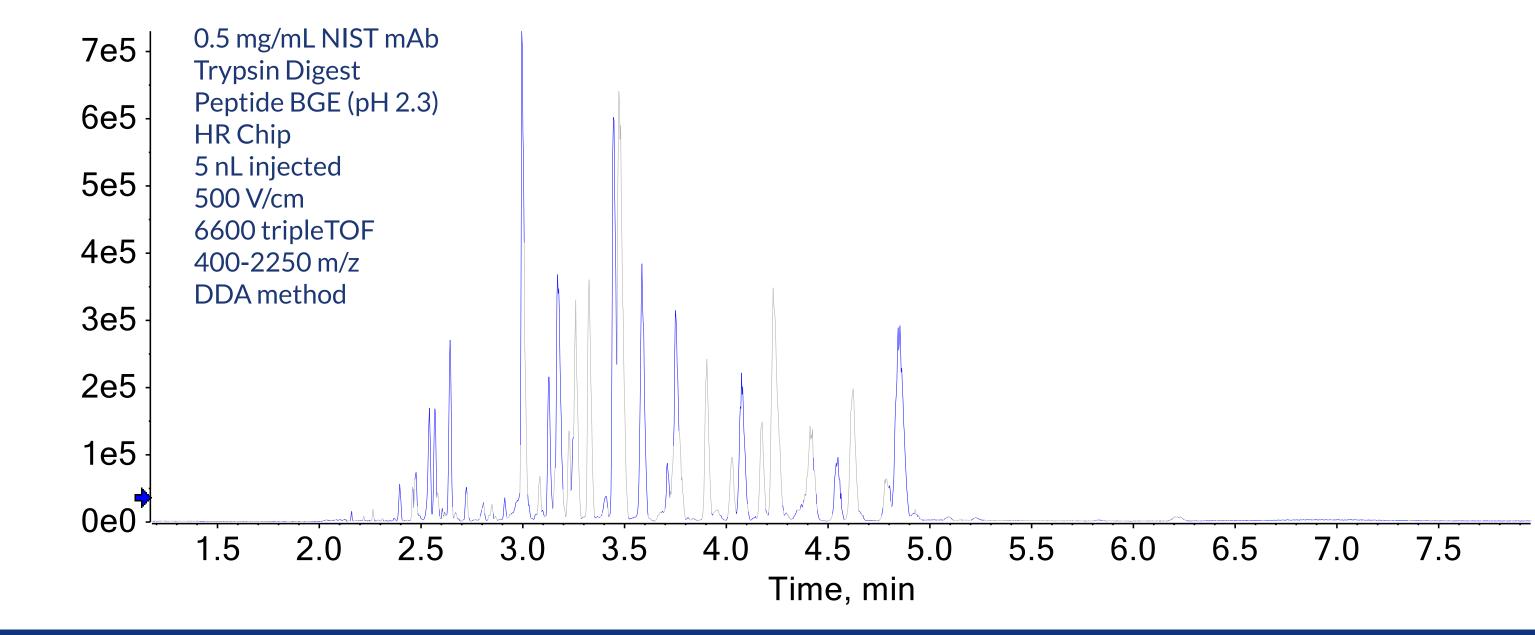
×908 devices

Intact Denatured Antibody 12000 6.5e4 11000 0.1 mg/mL NIST mAb 10000 9e6 **Reduced and IdeS** 0.5 mg/mL NIST mAb 6.0e4 9000 digested Peptide BGE (pH 2.3) 8000 Peptide BGE (pH 2.3) HR Chip 7000 5.5e4 8e6 6000[°] HR Chip 1 nL injected 5000 1 nL injected 500 V/cm 5.0e4 4000 500 V/cm 7e6 6600 tripleTOF 3000 6600 tripleTOF 1000 - 4000 m/z 2000 4.5e4 1000 - 3000 m/z 1000 6e6 3250 3150 3200 3300 4.0e4 Mass/Charge, Da Intensity 5e6 3.5e4 11000 sity 10000 9000 4e6 3.0e4 8000 7000 2.5e4 3e6 6000 5000 2.0e4 4000 2e6 3000 1.5e4 2000 1000 1e6 1.0e4 2400 2600 2800 3000 3200 3400 3600 3800 Mass/Charge, Da 0e0 5.0e3 2.0 2.5 Time, min

Antibody Subunits



Peptide Mapping



Summary and Acknowledgments

0.0e0

2.0

1.9

2.1

The work shown here demonstrates that the ZipChip methods developed for biopharmaceutical characterization can achieve excellent results when paired with the Sciex 6600 TripleTOF. The charge variant separation of the NIST mAb achieved baseline resolution and clear MS identification of the basic C-terminal lysine variants and deamidated acidic variants. The same method applied to 4 other biotherapeutic mAbs yielded excellent charge variant separations which closely match the well known charge variant characteristics of these molecules. The multilevel characterization performed in denaturing conditions demonstrates the versatility of methods that can be performed on the ZipChip system.

We thank Fang Wang and Zuzana Demianova of Sciex for hosting us in their lab, helping us to collect the data shown here, and providing the Humira, Infliximab, Rituximab, and Trastuzumab samples.

> The technologies discussed in this poster are the subject of one or more granted/pending patents. www.908devices.com/patents/